

Practical Use of AlphaFold and AlphaFold-Multimer

Mitch Miller

Rice University, Dec. 7, 2022

Overview

- Ways to obtain AlphaFold predictions
- How to assess AlphaFold predictions
- Example: running AlphaFold on Google Colab
- Some practical examples
 - Multimer predictions – finding interacting proteins
 - Ligands and Binding sites
 - Construct design
- Servers to help interpret the results

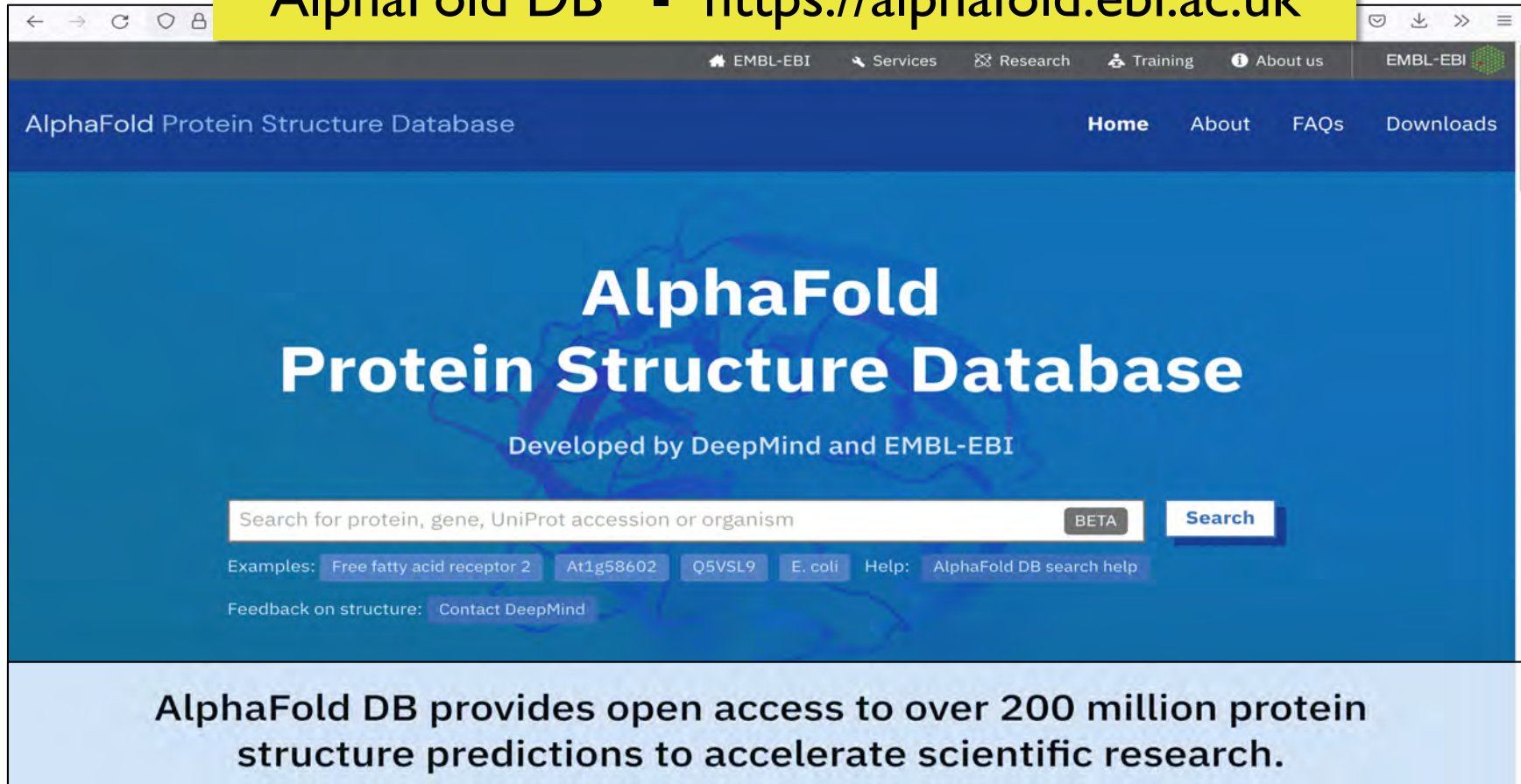
Ways to Obtain AlphaFold Predictions

- AlphaFoldDB
- AlphaFold via Google Colab
- Computer cluster
- Install locally

[Highly accurate protein structure prediction with AlphaFold.](#)

Jumper J, Evans R, Pritzel A, *et al.* 2021 *Nature* 596:583-589.

AlphaFold DB - <https://alphafold.ebi.ac.uk>



AlphaFold Protein Structure Database

Home About FAQs Downloads

AlphaFold Protein Structure Database

Developed by DeepMind and EMBL-EBI

Search for protein, gene, UniProt accession or organism BETA Search

Examples: [Free fatty acid receptor 2](#) [At1g58602](#) [Q5VSL9](#) [E. coli](#) Help: [AlphaFold DB search help](#)

Feedback on structure: [Contact DeepMind](#)

AlphaFold DB provides open access to over 200 million protein structure predictions to accelerate scientific research.

Tunyasuvunakool et al., (2021) Nature 596: 590–596

Search for protein, gene, UniProt accession or organism

BETA

Search

Examples: [Free fatty acid receptor 2](#)

[At1g58602](#)

[Q5VSL9](#)

[E. coli](#)

Help: [AlphaFold DB search help](#)

Tautomycetin biosynthetic PKS

AlphaFold structure prediction

Download

[PDB file](#)

[mmCIF file](#)

[Predicted aligned error](#)

Information

Protein	Tautomycetin biosynthetic PKS
Gene	I547_1222
Source organism	Mycobacterium kansasii 824 go to search
UniProt	X7YHF0 go to UniProt
Experimental structures	None available in the PDB
Biological function	Not available. go to UniProt

3D viewer

Model Confidence:

- Very high (pLDDT > 90)
- Confident (90 > pLDDT > 70)
- Low (70 > pLDDT > 50)
- Very low (pLDDT < 50)

Sequence of AF-X7YHF0-F1 Chain 1: Tautomyce... A

```
1  MTATLARLHVHGQSPAWPALYPHARQVELPTYPFQHHRYWLTFRSGTDAAGLGLDQPQHPLLGAIITTLADRDEVIATGRVVLGSAWLAHRVGDVVVLPATGFVDLVLGVGDYVGC
121  131  141  151  161  171  181  195  201  211  221  231
1  IDELVLHTPLVLAEHTPTDIQISVADADGARRSVNVHARTGTDHRAEAGWVLHASGTLRPGQTAAGEPPVPALPATAVDVQRFYQQLADCGLHYDPPFCVSRGIGLQPGDPDHIYAEV
241  251  261  271  281  291  301  311  321  331  341  351
1  ALPAGTDITGYGMHPALDDAALHPAAALGAGADAEPAVLRLEPFVFSGVTLYATAATRLQVWLTTRTGDDTFTLYASDPAGAPVIRIDTVVVRVLPDMATLSAPTTAAPSVPGLWELAWP
```



AlphaFold's Assessment of Model Quality

3D viewer

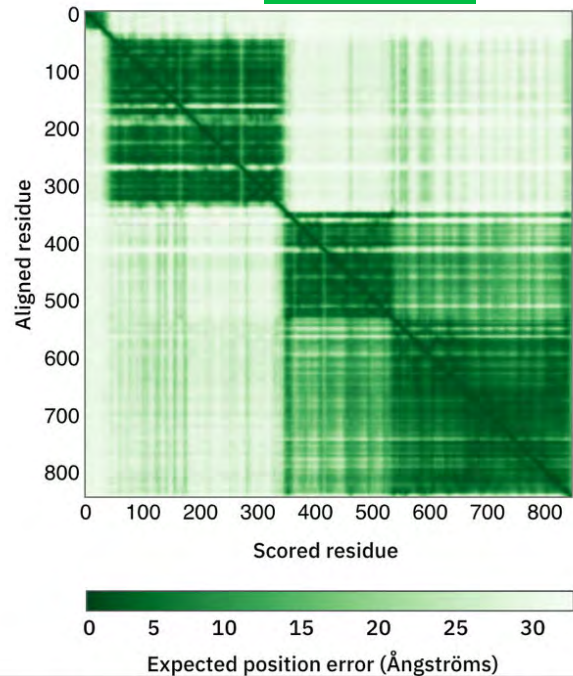
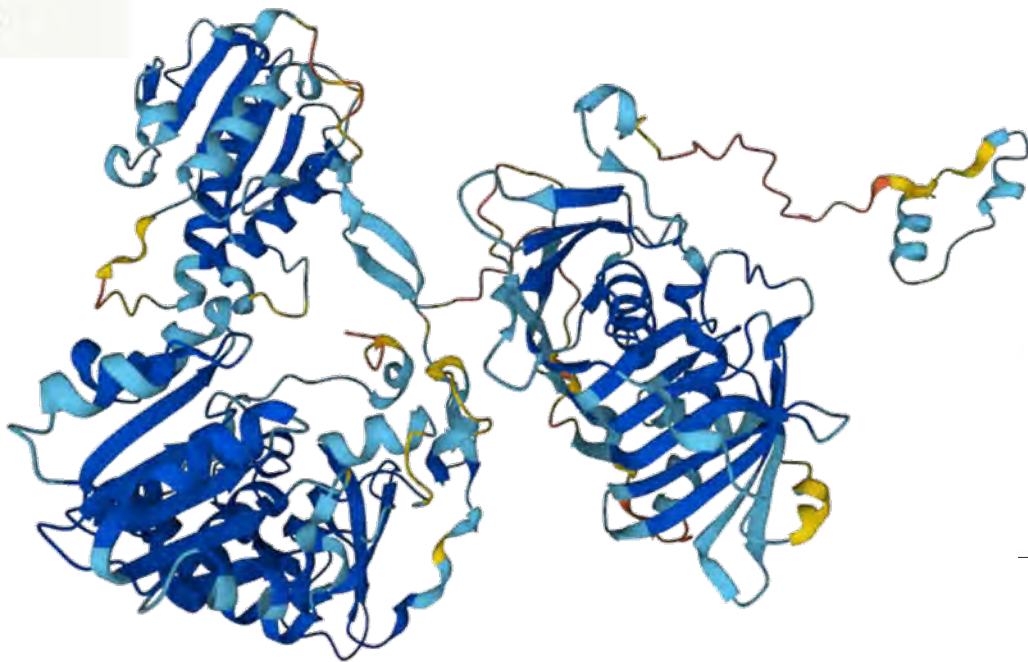
Model Confidence:

- Very high (pLDDT > 90)
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- Low (70 > pLDDT > 50)
- Very low (pLDDT < 50)

```
Sequence of AF-X7YHF0-F1 Chain 1: Tautomyce... A
1  MTATLARLHVHGQSPAWPALYPHARQVELPTYPFQHHRYWLTTPRSQTDAAAGLGDQPQHPHLLGAIITTLADRDEVIATGRVVLGSOAWLAHRVGDVVVLPATGFVDLVLGVGDYVGCVP
121  IDEVLVHTPLVLAHTPTDIQISVADAGAGRRSVNVHARTGTDHRAEAGWLVHASGTLRPGQTAAAGEPPVPALPATAVDVQRFYQQLADCGLHYDPPFCSVRGIGLQPGDHPHIYAEV
241  ALPAGTDITGYGMHPALLDAAALHPIAAALGAGADAEPAVLRLEPFVFSGVTLYATAATRQVWLTTRTGDDTFTLYASDPAGAPVIRIDTVVVRVLPDMATLSAPTTAAPSVEPLWELAWP
```

PAE

predicted Local Distance Difference Test



Predicted Aligned Error



bioRxiv

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New Results

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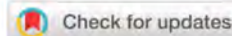
Protein complex prediction with AlphaFold-Multimer

Posted March 10, 2022

 Richard Evans,  Michael O'Neill,  Alexander Pritzel, Natasha Antropova,  Andrew Senior,  Tim Green, Augustin Žídek,  Russ Bates,  Sam Blackwell,  Jason Yim,  Olaf Ronneberger,  Sebastian Bodenstern, Michal Zielinski, Alex Bridgland,  Anna Potapenko,  Andrew Cowie,  Kathryn Tunyasuvunakool,  Rishub Jain,  Ellen Clancy,  Pushmeet Kohli,  John Jumper,  Demis Hassabis

doi: <https://doi.org/10.1101/2021.10.04.463034>

This article is a preprint and has not been certified by peer review [what does this mean?].



OPEN

ColabFold: making protein folding accessible to all

Milot Mirdita ^{1,10} , Konstantin Schütze ², Yoshitaka Moriwaki ^{3,4}, Lim Heo ⁵,
Sergey Ovchinnikov ^{6,7,10}  and Martin Steinegger ^{2,8,9,10} 

ColabFold offers accelerated prediction of protein structures and complexes by combining the fast homology search of MMseqs2 with AlphaFold2 or RoseTTAFold. ColabFold's 40–60-fold faster search and optimized model utilization enables prediction of close to 1,000 structures per day on a server with one graphics processing unit. Coupled with Google Colaboratory, ColabFold becomes a free and accessible platform for protein folding. ColabFold is open-source software available at <https://github.com/sokrypton/ColabFold> and its novel environmental databases are available at <https://colab-fold.mmseqs.com>.

Predicting the three-dimensional (3D) structure of a protein from its sequence alone remains an unsolved problem. However,

protein sizes of ~1,000 residues. For these, however, the MSA generation dominates the overall run time.

To enable researchers without these resources to use AlphaFold2, independent solutions based on Google Colaboratory were developed. Colaboratory is a proprietary version of Jupyter Notebook hosted by Google. It is accessible for free to logged-in users and includes access to powerful GPUs. Concurrently, Tunyasuvunakool et al.⁹ developed an AlphaFold2 Jupyter Notebook for Google Colaboratory (referred to as AlphaFold-Colab), for which the input MSA is built by searching with HMMer against the UniProt Reference Clusters (UniRef90) and an eightfold-reduced environmental database. This results in less accurate predictions while still requiring long search times.



colabfold



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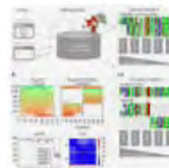
Tools

About 13,500 results (0.31 seconds)

<https://www.nature.com> > ... > brief communications

ColabFold: making protein folding accessible to all - Nature

by M Mirdita · 2022 · Cited by 805 — Here, we present **ColabFold**, a fast and easy-to-use software for the prediction of protein structures and homo- and heteromer complexes, for use ...



<https://github.com> > sokrypton > ColabFold

GitHub - sokrypton/ColabFold: Making Protein folding ...

LocalColabFold is an installer script designed to make **ColabFold** functionality available on local users' machines. It supports wide range of operating systems, ...

[AlphaFold2.ipynb](#) · [Issues 137](#) · [AlphaFold2_complexes.ipynb](#) · [README.md](#)



<https://colabfold.mmseqs.com>

ColabFold

ColabFold databases are MMseqs2 expandable profile databases to generate diverse multiple



🔗 main ▾

🔗 3 branches

📦 4 tags

Code ▾

About

Making Protein folding accessible to all!

📖 Readme

📄 MIT license

☆ 998 stars

👁 34 watching

🍴 271 forks

Releases 3

📦 v1.3.0 Latest
on Mar 4

+ 2 releases

Packages

No packages published



milot-mirdita Retry API requests a couple times if non-timeout errors...

5cadd35 4 days ago

🕒 1,139 commits

📁 .github

📁 MsaServer

📁 TemplateServer

📁 batch

📁 beta

📁 colabfold

📁 test-data

📁 tests

📁 utils

📁 verbose

📄 .gitattributes

☰ README.md



Making Protein folding accessible to all via Google Colab!

Notebooks	monomers	complexes	mmseqs2	jackhmmer	temp
AlphaFold2_mmseqs2	Yes	Yes	Yes	No	Yes
AlphaFold2_batch	Yes	Yes	Yes	No	Yes
RoseTTAFold	Yes	No	Yes	No	No
AlphaFold2 (from Deepmind)	Yes	Yes	No	Yes	No
ESMFold	Yes	Maybe	No	No	No
BETA (in development) notebooks					
AlphaFold2_advanced	Yes	Yes	Yes	Yes	No
OmegaFold	Yes	Maybe	No	No	No



ColabFold: AlphaFold2 using MMseqs2

Easy to use protein structure and complex prediction using [AlphaFold2](#) and [Alphafold2-multimer](#). Sequence alignments/templates are generated through [MMseqs2](#) and [HHsearch](#). For more details, see [bottom](#) of the notebook, checkout the [ColabFold GitHub](#) and read our manuscript. Old versions: [v1.0](#), [v1.1](#), [v1.2](#), [v1.3](#)

[Mirdita M, Schütze K, Moriwaki Y, Heo L, Ovchinnikov S, Steinegger M. ColabFold: Making protein folding accessible to all. Nature Methods, 2022](#)



▶ Input protein sequence(s), then hit Runtime -> Run all

🎧 `query_sequence: "PIAQIHILEGRSDEQKETLIREVSEAI SRSLDAPLTSVRVIITEMAKGHFGIGGELASK"`

- Use `:` to specify inter-protein chainbreaks for **modeling complexes** (supports homo- and hetro-oligomers). For example `PI...SK:PI...SK` for a homodimer

`jobname: "test"`

`use_amber:`

`template_mode: none` ▾

- "none" = no template information is used, "pdb70" = detect templates in pdb70, "custom" - upload and search own templates (PDB or mmCIF format, see [notes below](#))



Easy to use protein structure and complex prediction using [AlphaFold2](#) and [Alphafold2-multimer](#). Sequence alignments/templates are generated through [MMseqs2](#) and [HHsearch](#). For more details, see [bottom](#) of the notebook, checkout the [ColabFold GitHub](#) and read our manuscript. Old versions: [v1.0](#), [v1.1](#), [v1.2](#), [v1.3](#)

[Mirdita M, Schütze K, Moriwaki Y, Heo L, Ovchinnikov S, Steinegger M. ColabFold: Making protein folding accessible to all. Nature Methods, 2022](#)



▶ Input protein sequence(s), then hit Runtime -> Run all

▶ **query_sequence:** "FH PNVHFKTGEICLDILKNAWSPAATLQSVCRRAIALMAHPEPDSPLNCDSGNLLRSGDVRGFNSMAQMYTRLAAMPKKGLE VLFQGP"

- Use **:** to specify inter-protein chainbreaks for **modeling complexes** (supports homo- and hetro-oligomers). For example **PI...SK:PI...SK** for a homodimer

jobname: "PEX4"

use_amber:

template_mode: none

- "none" = no template information is used, "pdb70" = detect templates in pdb70, "custom" - upload and search own templates (PDB or mmCIF format, see [notes below](#))

[Show code](#)



▶ MSA options (custom MSA upload, single sequence, pairing mode)

msa_mode: MMseqs2 (UniRef+Environmental)

pair_mode: unpaired+paired

- "unpaired+paired" = pair sequences from same species + unpaired MSA, "unpaired" = separate MSA for each chain, "paired" - only use paired sequences.

[Show code](#)

▶ Advanced settings

model_type: auto

- "auto" = protein structure prediction using "AlphaFold2-ptm" and complex prediction "AlphaFold-multimer-v2". For complexes "AlphaFold-multimer-v[1,2]" and "AlphaFold-ptm" can be used.

num_recycles: 3

save_to_google_drive:

- if the save_to_google_drive option was selected, the result zip will be uploaded to your Google Drive

dpi: 200

- set dpi for image resolution



Show code

Instructions

Quick start

1. Paste your protein sequence(s) in the input field.
2. Press "Runtime" -> "Run all".
3. The pipeline consists of 5 steps. The currently running step is indicated by a circle with a stop sign next to it.

Result zip file contents

1. PDB formatted structures sorted by avg. pLDDT and complexes are sorted by pTMScore. (unrelaxed and relaxed if use_amber is enabled).
2. Plots of the model quality.
3. Plots of the MSA coverage.
4. Parameter log file.
5. A3M formatted input MSA.
6. A predicted_aligned_error_v1.json using [AlphaFold-DB's format](#) and a scores.json for each model which contains an array (list of lists) for PAE, a list with the average pLDDT and the pTMScore.
7. BibTeX file with citations for all used tools and databases.

At the end of the job a download modal box will pop up with a `jobname.result.zip` file. Additionally, if the `save_to_google_drive` option was selected, the `jobname.result.zip` will be uploaded to your Google Drive.



+ Code + Text



Run all ⌘/Ctrl+F9

Run before ⌘/Ctrl+F8

Run the focused cell ⌘/Ctrl+Enter

Run selection ⌘/Ctrl+Shift+Enter

Run after ⌘/Ctrl+F10

Interrupt execution ⌘/Ctrl+M

Restart runtime ⌘/Ctrl+M

Restart and run all

Disconnect and delete runtime

Change runtime type

Manage sessions

View runtime logs

Connect Editing



ColabFold: Alpha

Easy to use protein structure alignments/templates notebook, checkout the [Mirdita M, Schütze K, N accessible to all. Nature](#)

Input protein sequ



query_seque

[d2](#) and [AlphaFold2-multimer](#). Sequence [ch](#). For more details, see [bottom](#) of the Old versions: [v1.0](#), [v1.1](#), [v1.2](#), [v1.3](#) [I. ColabFold: Making protein folding](#)



```
QSVCRAIIALMAHPEPDSPLNCDSGNLLRSGDVRGFNSMAQMYTRLAAMPKKGLE VLFQGP "
```

- Use `:` to specify inter-protein chainbreaks for **modeling complexes** (supports homo- and hetro-oligomers). For example `PI...SK:PI...SK` for a homodimer

jobname: " PEX4 "

use_amber:

template_mode: none

- "none" = no template information is used, "pdb70" = detect templates in pdb70, "custom" - upload and search own templates (PDB or mmCIF format, see [notes below](#))





ColabFold: AlphaFold2 using MMseqs2

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[Mirdita M, Schütze K, Moriwaki Y](#)
[accessible to all. Nature Methods](#)



Warning: This notebook was not authored by Google.

This notebook is being loaded from [GitHub](#). It may request access to your data stored with Google, or read data and credentials from other sessions. Please review the source code before executing this notebook.

Cancel [Run anyway](#)

Input protein sequence(s)

query_sequence: "M..."

- Use : to specify inter-domain boundaries for homodimer

jobname: "PEX4"

use_amber:

template_mode: none

- "none" = no template information is used, "pdb70" = detect templates in pdb70, "custom" - upload and search own templates (PDB or mmCIF format, see [notes below](#))

...QPPQVRFLTKIFH PNVHFKTGEICLD "

For example `PI...SK:PI...SK` for a

⚠️ Don't forget to hit Runtime -> Run all after updating the form.

[Show code](#)

▶ Install dependencies

✓ [4] [Show code](#)

▶ Run Prediction

🔍 [Show code](#)

```
/usr/local/lib/python3.8/dist-packages/Bio/Data/SCOPData.py:18: BiopythonDeprecationWarning: The 'Bio.Data.SCOPData' module will
warnings.warn(
Downloading alphafold2 weights to .: 100%|██████████| 3.47G/3.47G [01:48<00:00, 34.3MB/s]
2022-12-06 03:39:21,053 Found 5 citations for tools or databases
2022-12-06 03:39:21,073 Unable to initialize backend 'tpu_driver': NOT_FOUND: Unable to find driver in registry given worker:
2022-12-06 03:39:21,819 Unable to initialize backend 'rocm': NOT_FOUND: Could not find registered platform with name: "rocm". Av
2022-12-06 03:39:21,820 Unable to initialize backend 'tpu': module 'jaxlib.xla_extension' has no attribute 'get_tpu_client'
2022-12-06 03:39:21,821 Unable to initialize backend 'plugin': xla_extension has no attributes named get_plugin_device_client. C
2022-12-06 03:39:27,786 Query 1/1: PEX4_71d3d (length 165)
PENDING: 0% | 0/150 [elapsed: 00:00 remaining: ?]2022-12-06 03:39:28,608 Sleeping for 9s. Reason: PENDING
RUNNING: 6% || 9/150 [elapsed: 00:10 remaining: 02:46]2022-12-06 03:39:38,425 Sleeping for 10s. Reason: RUNNING
```

▶ Display 3D structure

🔍 rank_num: 1



+ Code + Text 📄 Copy to Drive

RAM Disk Editing ⏴

Show code

```
/usr/local/lib/python3.8/dist-packages/Bio/Data/SCOPData.py:18: BiopythonDeprecationWarning: The 'Bio.Data.SCOPData' module will
warnings.warn(
Downloading alphafold2 weights to .: 100%|██████████| 3.47G/3.47G [01:48<00:00, 34.3MB/s]
2022-12-06 03:39:21,053 Found 5 citations for tools or databases
2022-12-06 03:39:21,073 Unable to initialize backend 'tpu_driver': NOT_FOUND: Unable to find driver in registry given worker:
2022-12-06 03:39:21,819 Unable to initialize backend 'rocm': NOT_FOUND: Could not find registered platform with name: "rocm". Av
2022-12-06 03:39:21,820 Unable to initialize backend 'tpu': module 'jaxlib.xla_extension' has no attribute 'get_tpu_client'
2022-12-06 03:39:21,821 Unable to initialize backend 'plugin': xla_extension has no attributes named get_plugin_device_client. C
2022-12-06 03:39:27,786 Query 1/1: PEX4_71d3d (length 165)
PENDING: 0%|          | 0/150 [elapsed: 00:00 remaining: ?]2022-12-06 03:39:28,608 Sleeping for 9s. Reason: PENDING
RUNNING: 6%|█        | 9/150 [elapsed: 00:10 remaining: 02:46]2022-12-06 03:39:38,425 Sleeping for 10s. Reason: RUNNING
RUNNING: 13%|██       | 19/150 [elapsed: 00:21 remaining: 02:26]2022-12-06 03:39:49,230 Sleeping for 5s. Reason: RUNNING
COMPLETE: 100%|██████| 150/150 [elapsed: 00:29 remaining: 00:00]
2022-12-06 03:39:58,230 Running model_1
2022-12-06 03:41:32,133 model_1 took 88.5s (3 recycles) with pLDDT 90.2 and ptmscore 0.855
```

colored by N→C

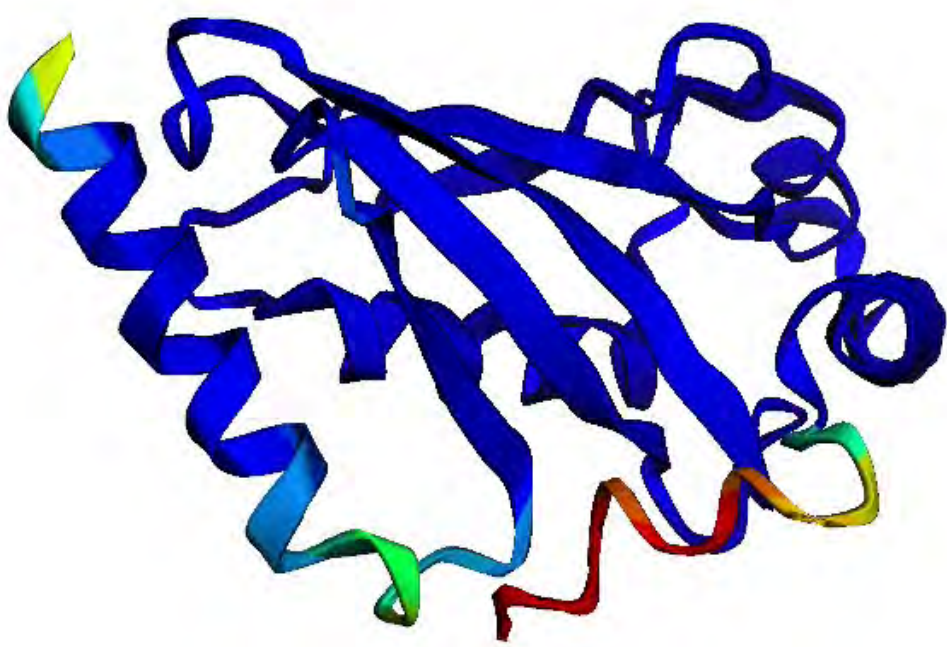


colored by pLDDT

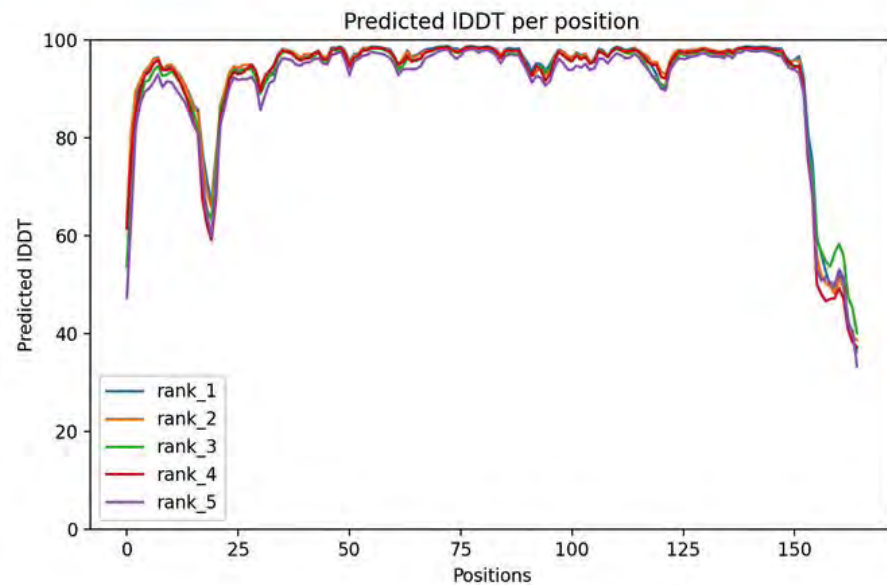
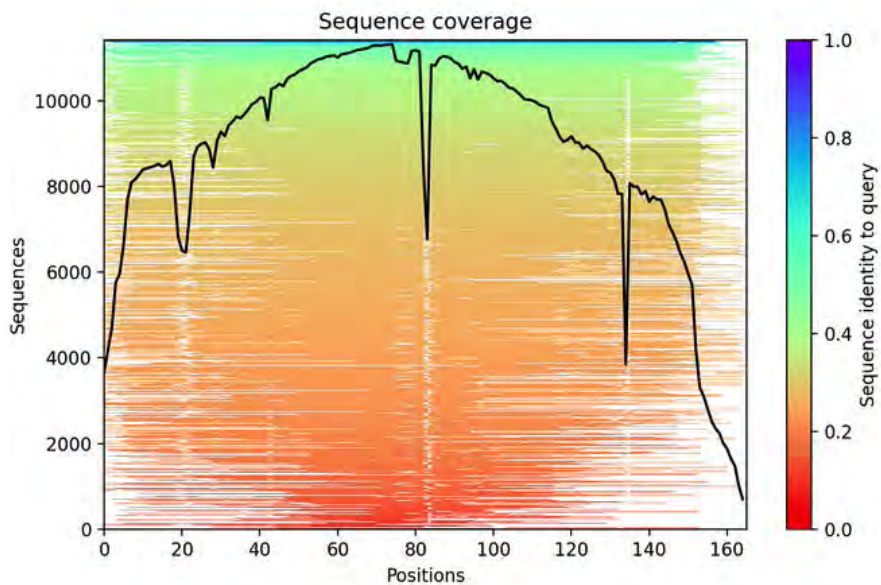
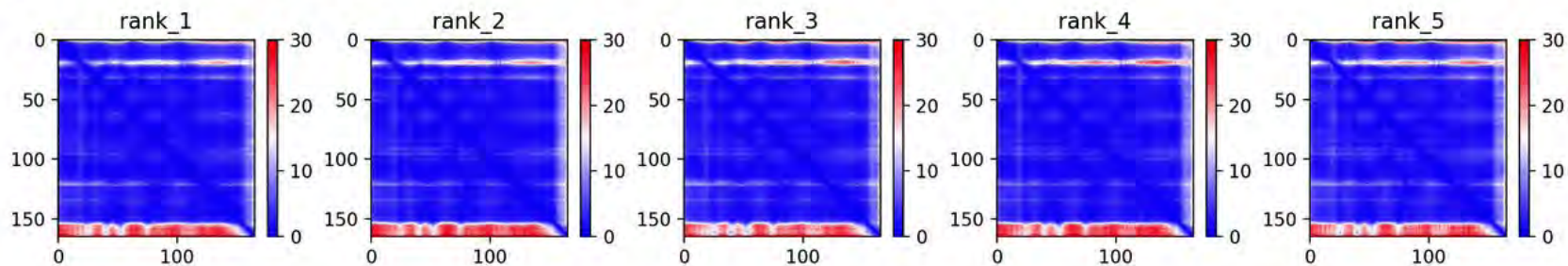




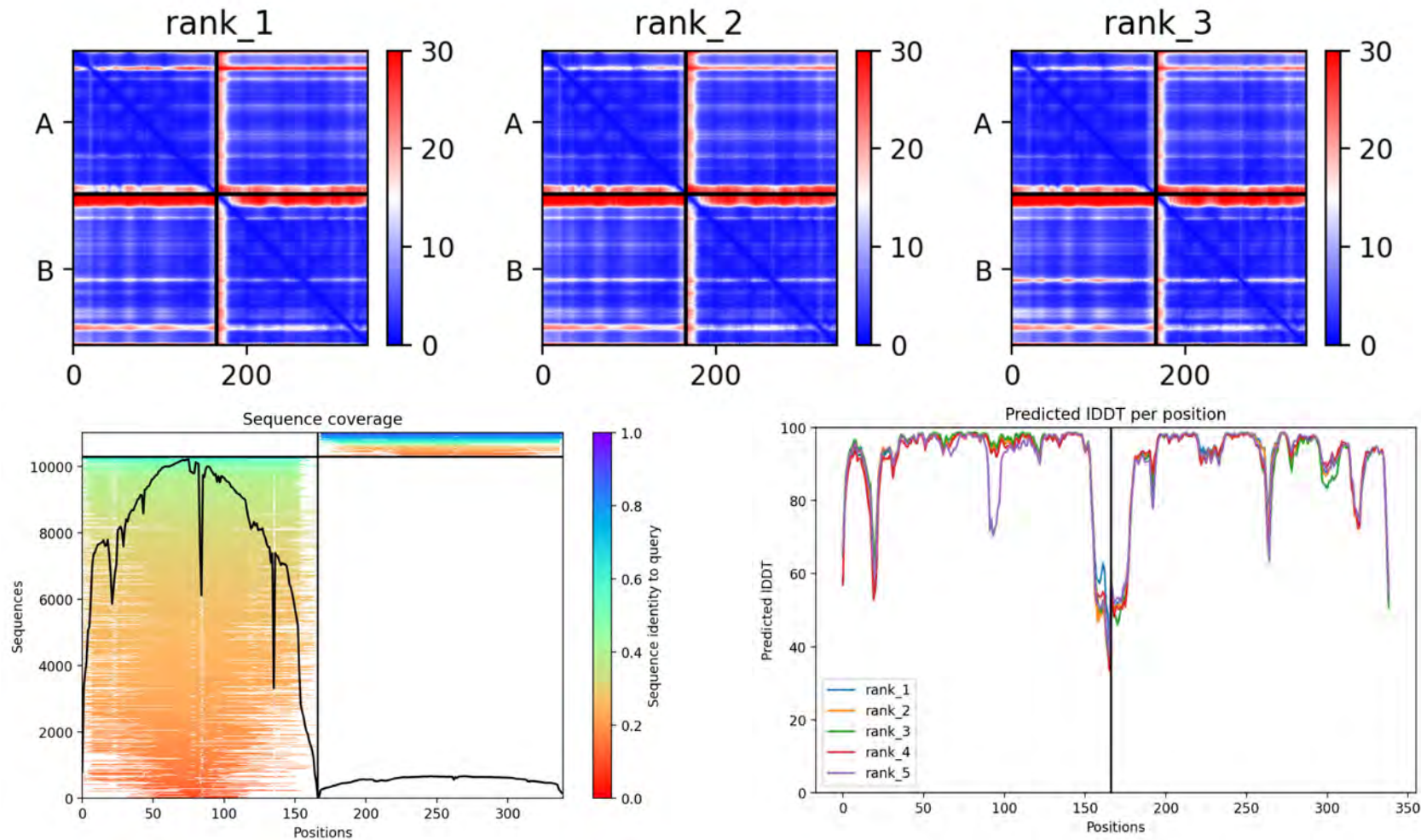
```
0s ✓ ▶ show_sidechains:   
  
show_mainchains:   
  
Show code
```



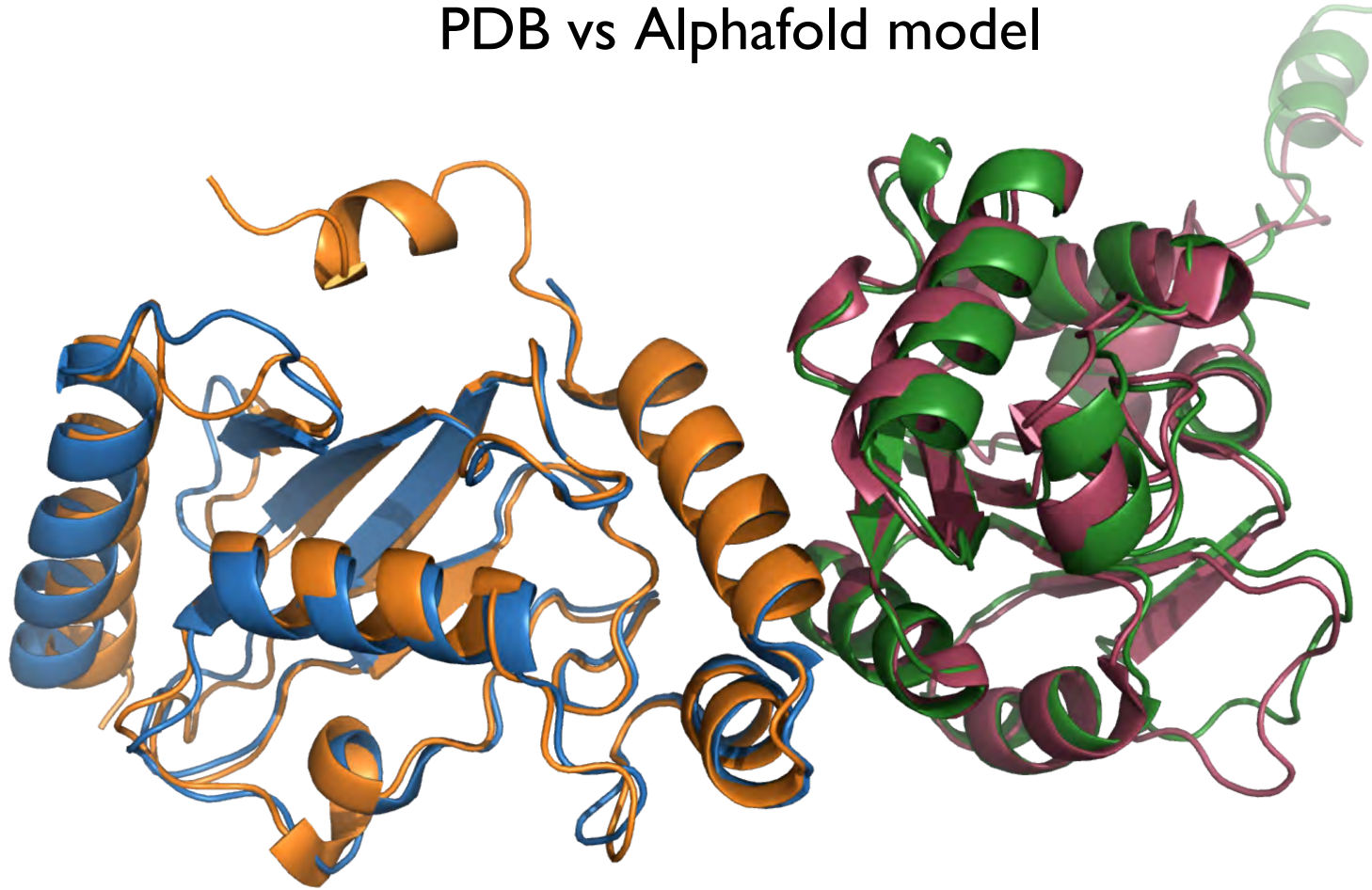
Plots for PEX4_71d3d



PEX4:PEX22 heterodimer



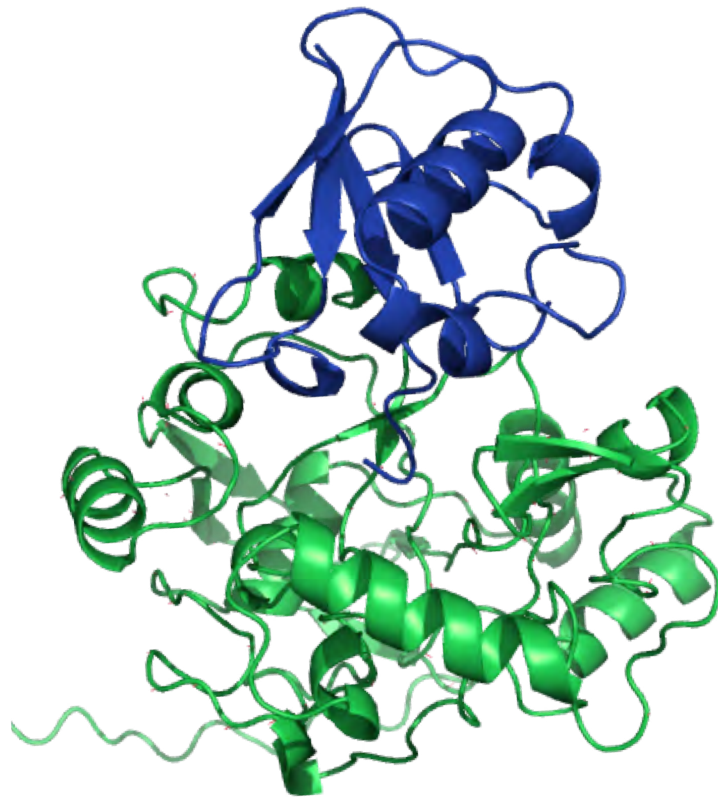
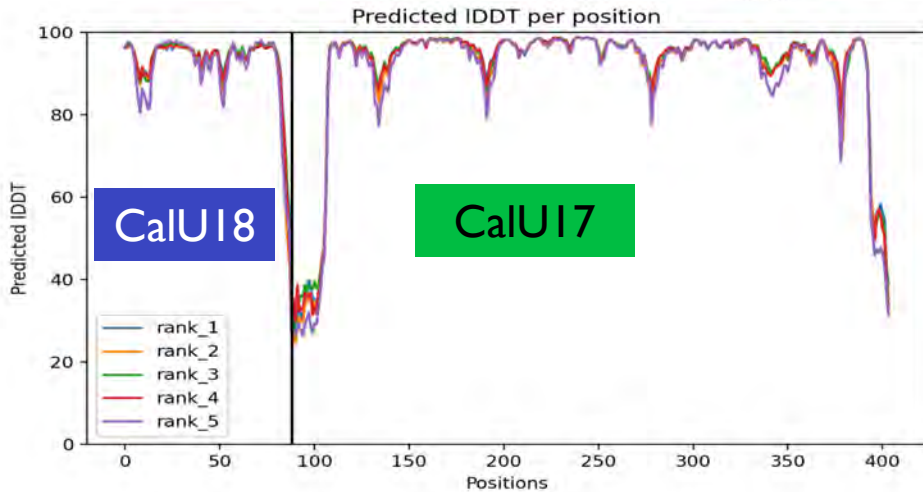
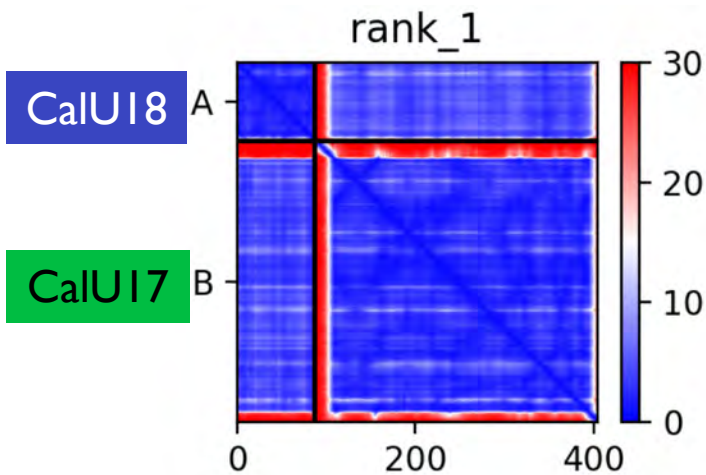
PEX4:PEX22 heterodimer PDB vs AlphaFold model



CaUI7 CaUI8 heterodimer prediction



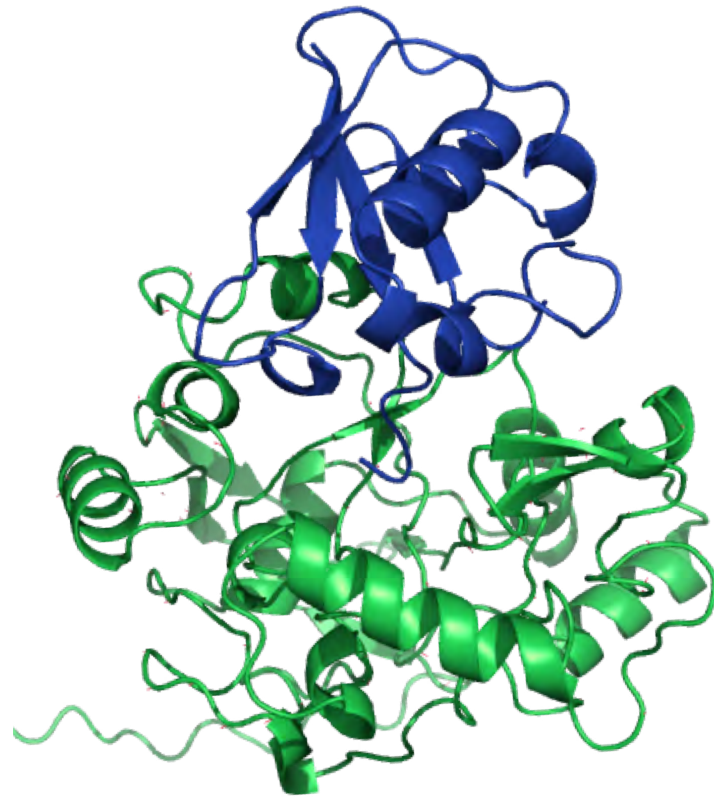
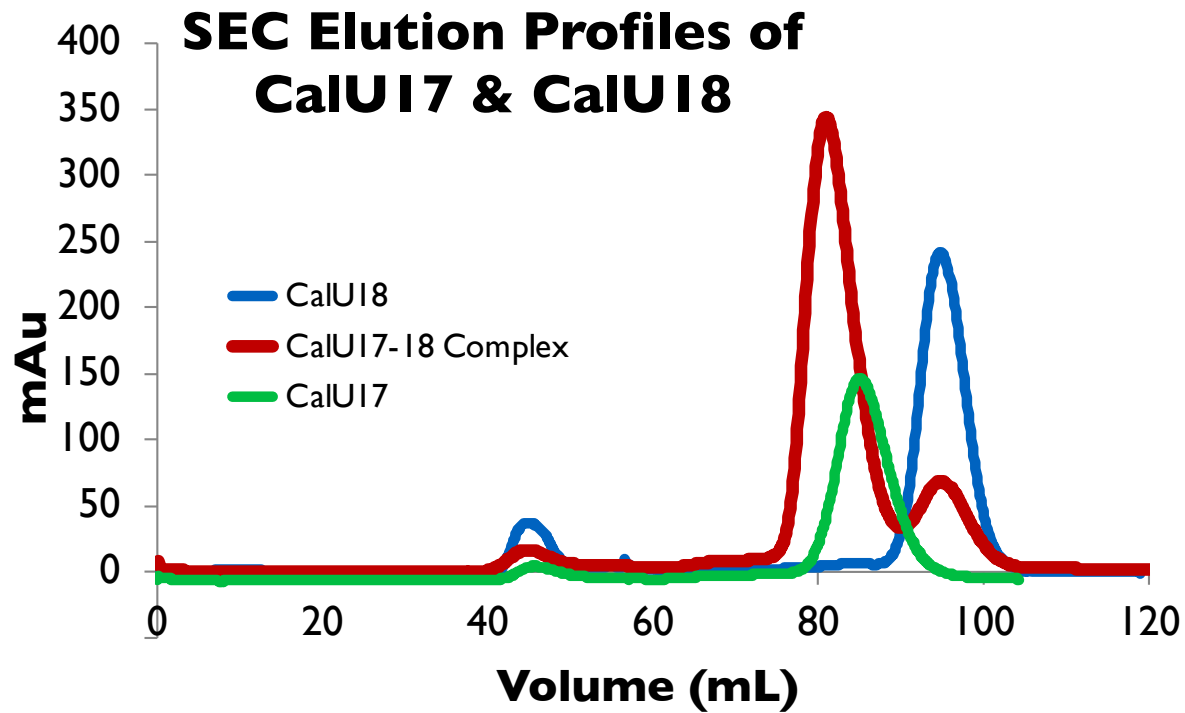
Abigael
Kosgei



CaUI7 CaUI8 heterodimer prediction



Abigael
Kosgei





AlphaFill

AlphaFold models enriched with ligands and co-factors

Uniprot ID

Browse...

AlphaFill

AlphaFill is an algorithm based on sequence and structure similarity that “transplants” missing ligands, cofactors and (metal) ions to the AlphaFold models. By adding the molecular context to these protein structures, the models can be more easily appreciated in terms of function and structural integrity. Consequently, the AlphaFill models can be helpful in designing downstream wet-lab experiments and/or computational studies.

AlphaFill databank

The AlphaFill databank contains all AlphaFold models including the transferred compounds, if present. For each entry, metadata is provided describing the origin of the ligands and indicating the quality based on global and local Root-Mean-Square deviation (RMSd) values.

Data for a single entry can be found at the corresponding entry page (search by UniProt ID above). Download details for the complete databank can be found at the [download page](#).

For more AlphaFill details: <https://www.nature.com/articles/s41592-022-01685-y>

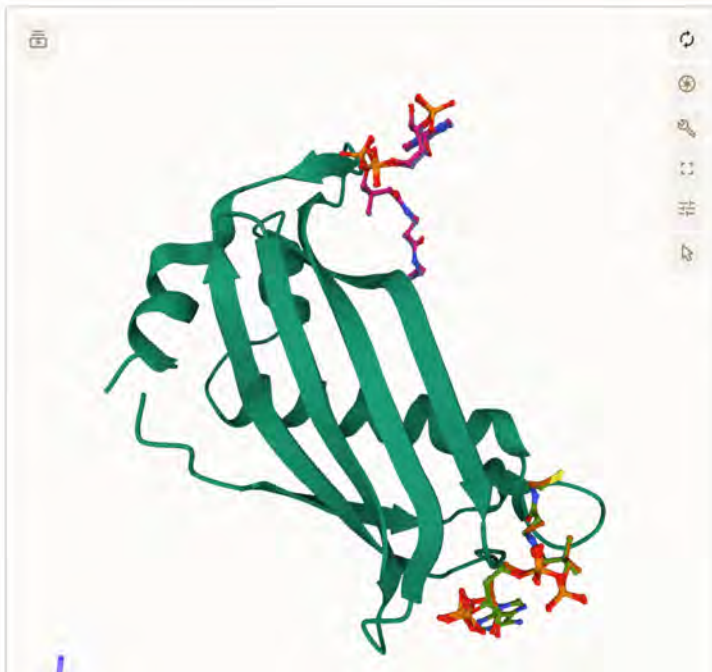
P0A8Z3

Acyl-CoA thioester hydrolase YbgC

Structure file <https://alphafill.eu/v1/aff/P0A8Z3-F1>

Metadata <https://alphafill.eu/v1/aff/P0A8Z3-F1/json>

Original AlphaFold model <https://alphafold.ebi.ac.uk/entry/P0A8Z3>



		25% identity	30% identity	40% identity	50% identity	60% identity	70% identity
		Global		Local			
Compound	PDBID	RMSd	Asym	RMSd	TCS	<input checked="" type="checkbox"/>	Show
COA	5kl9.A	0.51	C	0.13	0.08	<input checked="" type="checkbox"/>	
			B	0.16	0.07	<input checked="" type="checkbox"/>	
HXC ->	5t06.A	0.29	D	0.14	0.07	<input checked="" type="checkbox"/>	
COA	5t06.C	0.84	E	0.18	0.04	<input checked="" type="checkbox"/>	

<https://search.foldseek.com/search>

Foldseek Search



[GITHUB](#)

[SÖDING LAB](#)

[STEINEGGER LAB](#)

Queries



ATOM	1	N	MET	A	1	-23.382	11.235	10.663	1.00	61.85	N
ATOM	2	CA	MET	A	1	-22.302	10.835	11.561	1.00	61.85	C
ATOM	3	C	MET	A	1	-22.533	9.427	12.098	1.00	61.85	C
ATOM	4	CB	MET	A	1	-22.175	11.823	12.723	1.00	61.85	C
ATOM	5	O	MET	A	1	-21.591	8.642	12.220	1.00	61.85	O
ATOM	6	CG	MET	A	1	-20.797	11.840	13.365	1.00	61.85	C
ATOM	7	SD	MET	A	1	-20.701	12.988	14.793	1.00	61.85	S
ATOM	8	CE	MET	A	1	-20.491	14.566	13.920	1.00	61.85	C
ATOM	9	N	GLN	A	2	-23.731	9.029	12.273	1.00	81.56	N
ATOM	10	CA	GLN	A	2	-24.088	7.743	12.861	1.00	81.56	C
ATOM	11	C	GLN	A	2	-23.892	6.607	11.861	1.00	81.56	C
ATOM	12	CB	GLN	A	2	-25.535	7.761	13.357	1.00	81.56	C
ATOM	13	O	GLN	A	2	-23.436	5.522	12.228	1.00	81.56	O
ATOM	14	CG	GLN	A	2	-25.675	8.118	14.831	1.00	81.56	C
ATOM	15	CD	GLN	A	2	-27.119	8.130	15.297	1.00	81.56	C



Search Settings

Databases

- AlphaFold/UniProt50 v3
- AlphaFold/Swiss-Prot v2
- AlphaFold/Proteome v2
- MGnify-ESM30 v1
- PDB100 220722
- GMGCL 2204

Mode

- 3Di/AA
- TM-align

Taxonomic filter

[CURL COMMAND](#)

[LOAD ACCESSION](#)

[UPLOAD PDB](#)

[PREDICT STRUCTURE](#)

[SEARCH](#)

van Kempen et al., 2022. bioRxiv <https://doi.org/10.1101/2022.02.07.479398>

<https://search.foldseek.com/search>

Foldseek Search

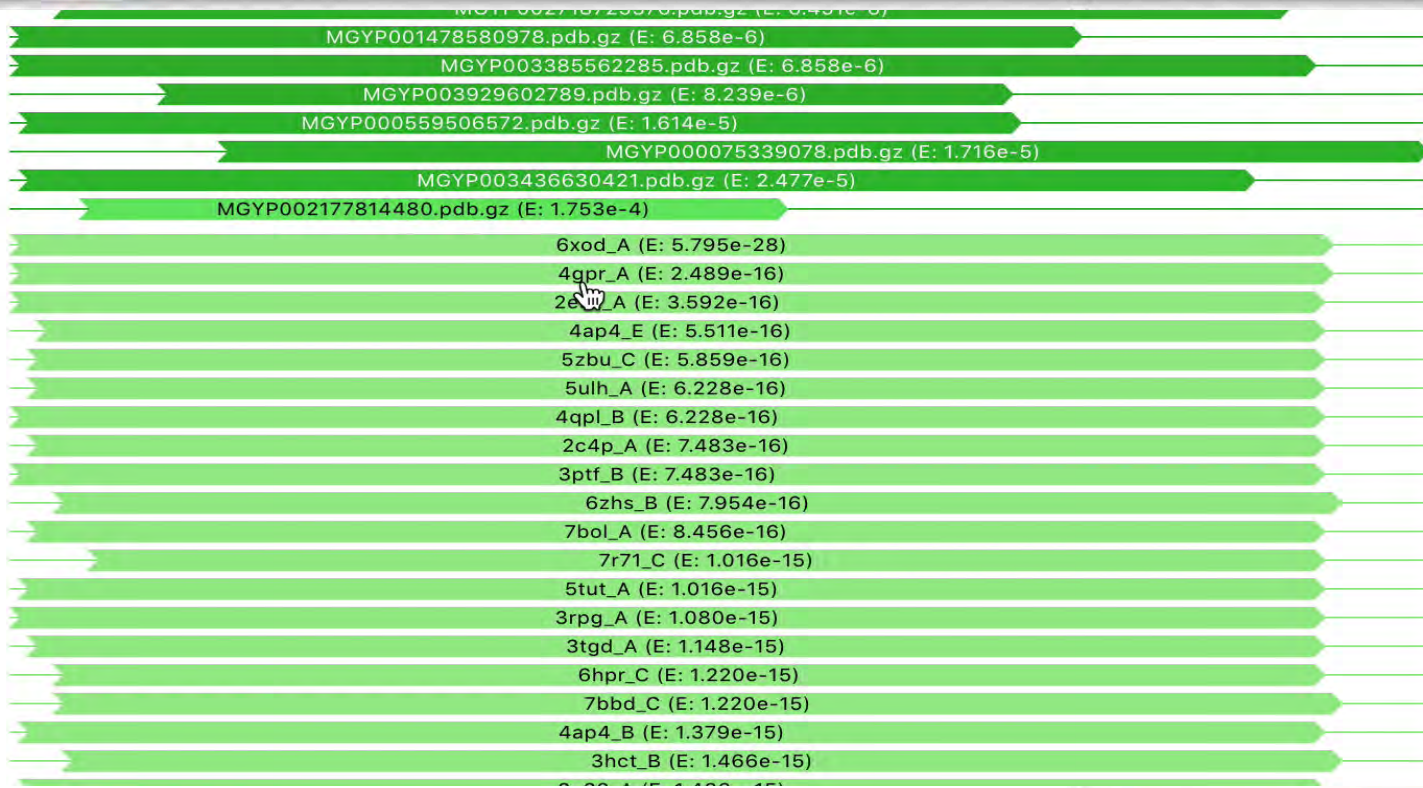


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van Kempen et al., 2022. bioRxiv <https://doi.org/10.1101/2022.02.07.479398>

<https://search.foldseek.com/search>

Foldseek Search



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Query Position

1-154 (165)

Target Position

1-150 (150)

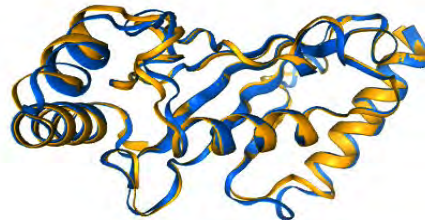


```
Q 1 MQASRARLFKEYKEVQREKVADP-DIQLICDDTNIFKWTALIKGPSETPYEGGVFQLAFSVPEPYPLQPPQVRFLTKIFH
   + R+ KE +E+Q++ P + +IF WTA I GP ++PY+GG+F L P YP++ P+V F+TK++H
T 1 AAMAMRRIQKELREIQD---PPCNC SAGPVGDDIFHW TATITGPDDSPYQGGLFFLDVHFPVDYPFKAPRVTFMTKVVH

Q 80 PNVHFKTGEICLDILKNAWSPA WTLQSVCR AIALMAHPEPDSPLNCD SGNLLRSGDVRGFNSMAQMYTRLAAMP
   PN++ G ICLDILK+ WSPA TL V +I +L+ P P PL+ + +N+LR+ + + F A+ +TR+ A P
T 78 PNINK-NGVICLDILKDQWSPAL T LSRVLLSISLLTDPNPSDPLDPEVANVLRA-NKKQFEDTAREWTRMYARP
```

TM-Score: 0.95185

Select target residues to highlight their structure



Conclusions

Exciting times

Easy to access structures

New creative ways of using structure predications

Predictions are great hypotheses

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George Phillips

Alex Ditzel

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David Xu

